



State of Oklahoma
Department of Agriculture, Food, and Forestry

Mary Fallin
Governor

Jim Reese
Secretary of Agriculture

April 15, 2014

Tawanda Maignan, Team Leader
Emergency Response Team
U.S.EPA Office of Pesticide Programs
Document Processing Desk (EMEX)
Room S4900, One Potomac Yard
2777 Crystal Drive
Arlington, VA 22202

Subject: Request for a Section 18 specific exemption for use of **Transform WG Insecticide**, EPA Registration Number 62719-625 to be applied on sorghum fields (grain & forage) to control the sugarcane aphid in Oklahoma.

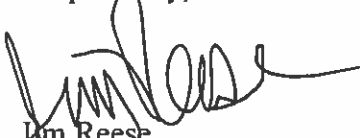
Tawanda Maignan:

The Oklahoma Department of Agriculture, Food, and Forestry (ODAFF) requests a specific emergency exemption under the provisions of section 18 of the Federal Insecticide Fungicide and Rodenticide Act, as amended, for the use of Transform WG Insecticide, EPA Registration Number 62719-625 to be applied on sorghum fields (grain & forage) to control the sugarcane aphid in Oklahoma.

This is the first year ODAFF has requested a specific emergency exemption for this use using this product. The Department asks that this specific exemption request be processed as an expedited request. The aphid has since been discovered in Louisiana and Texas. These states have indicated that they also want to pursue this Section 18. Documented economic damage from this pest ranges from 20% to 100% yield loss.

If you have any questions in connection with this petition, please contact Ryan Williams, (405) 522-5993. Thank you for your consideration of our exemption request.

Respectfully,


Jim Reese
Secretary of Agriculture
Enclosures



Oklahoma Sorghum Association

4201 North Interstate 27
Lubbock, Texas 79403
(800) 658-9808

February 13, 2014

Ryan Williams
Pesticide Certification & Training Administrator
Oklahoma Department of Agriculture Food and Forestry
2800 N. Lincoln Blvd
Oklahoma City, OK 73105

Dear Mr. Williams:

On behalf of the Oklahoma Sorghum Association I am writing to affirm our organizations' support for the Section 18 permit for *Transform*. This invasive pest has the potential to cause agronomic and economic harm to sorghum farmers in many different regions of the Sorghum Belt.

Currently there is no chemical labeled or cultural practice demonstrated to have significant effectiveness in controlling the aphid, therefore, we believe a Section 18 permit for the use of *Transform* as a pesticide infestation control during the 2014 growing season should be approved as soon as possible.

Both National Sorghum Producers and the United Sorghum Checkoff Program along with farmers and entomologist, including Oklahoma State Extension, have been working diligently to determine the best possible approach to provide sorghum farmers with a control method to manage this new pest.

Therefore, the Oklahoma Sorghum Association strongly supports timely approval of the Section 18 permit for *Transform*. Thank you for your consideration, and if you have any questions or concerns, please feel free to reach out to our organization.

Sincerely,

Jordan Shearer, President
Oklahoma Sorghum Association



February 13, 2014

Ryan Williams
Pesticide Certification & Training Administrator
Oklahoma Department of Agriculture Food and Forestry
2800 N. Lincoln Blvd
Oklahoma City, OK 73105

Dear Mr. Williams:

National Sorghum Producers supports the Section 18 permit for *Transform* to avoid agronomic and economic implications for sorghum farmers in the southern and eastern regions of the Sorghum Belt. The aphid identified as *Melanaphis Sacchari* has become an invasive pest, effecting thousands of U.S. sorghum acres. NSP feels decisive action is needed given that no chemical or cultural practice has demonstrated significant efficacy and asks that a Section 18 permit for the use of *Transform* as a pesticide to control infestations during the 2014 growing season be approved as soon as possible

The urgency NSP places on this request is vital to the success of our crop and the acres expected to be planted during the 2014 growing season. In fact, nearly 25 percent of the U.S. sorghum crop will begin to be planted as early as Feb. 1, 2014. Without a resource to control this new pest plaguing the southern and eastern regions of the Sorghum Belt, nearly 100 million bushels of sorghum with a potential market value of \$400 million could be affected.

The aphid has been identified in many key sorghum producing states, including Oklahoma. It is important a method of control for this invasive pest be implemented quickly to avoid devastating impacts to sorghum farmers, rural economies and the entire U.S. sorghum industry.

NSP has worked diligently with the United Sorghum Checkoff Program, farmers, entomologists and other involved parties on the best possible approach to provide sorghum farmers with a control method to manage this new pest. Therefore, NSP strongly supports timely approval of the Section 18 permit for *Transform*. NSP appreciates your attention to this matter and for your support. Please contact us if you have any questions or concerns.

Sincerely,

A handwritten signature in black ink that reads "Tim Lust".

Tim Lust, CEO
National Sorghum Producers



Committed to invest sorghum checkoff dollars efficiently to increase profitability for sorghum.

March 31, 2014

Ryan Williams
Pesticide Certification & Training Administrator
Oklahoma Department of Agriculture Food and Forestry
2800 N. Lincoln Blvd
Oklahoma City, OK 73105

Dear Mr. Williams:

As the national research, promotion, and education board for the U.S. sorghum industry, the United Sorghum Checkoff Program (USCP) is committed to addressing the needs of U.S. sorghum farmers. Pest management remains a major agronomic objective of the USCP and the recent developments regarding a new aphid impacting the southern and eastern Sorghum Belt merits precise and immediate attention.

Given the scope of this new aphid's range, the USCP is strongly supportive of efforts to approve the Section 18 permit for the use of *Transform* as a pesticide to control infestations during the 2014 growing season. If left uncontrolled in the southern and eastern Sorghum Belt, this new pest has the ability to impact nearly 100 million bushels of sorghum with a potential market value of \$400 million. Given that the aphid has been identified in multiple key sorghum states, including Oklahoma, the need for action is crucial and timely. Effectively providing control options to sorghum farmers potentially impacted by this pest is needed to avoid devastating implications to the U.S. sorghum industry.

The USCP is confident in the scientific and strategic approach documented by the agencies involved in this effort. The USCP strongly supports the timely approval of the Section 18 permit for *Transform*. This chemical will provide a documented control method for sorghum farmers to manage this new pest. We appreciate your support in this effort as well as the affiliated institutions who have been addressing this critical issue. Please contact me with any questions or concerns.

Justin Weinheimer, Ph.D.
United Sorghum Checkoff Program
Crop Improvement Program Director



Oklahoma Cooperative Extension Service
Division of Agricultural Sciences and Natural Resources
Oklahoma State University

Department of Entomology and Plant Pathology • 127/110 Noble Research Center
Stillwater, Oklahoma 74078-3033 • (405) 744-5527 • Fax (405) 744-6039
www.ento.okstate.edu

February 24, 2014

Ryan Williams
Oklahoma Department Of Ag., Food, & Forestry
Certification & Training Administrator
2800 N. Lincoln Blvd.
Oklahoma City, Ok 73105

Ryan:

We are requesting the section 18 for the use of sulfoxaflor to control a potentially injurious aphid, the sugarcane aphid *Melanaphis sacchari* which was found in Texas, Louisiana, and in Bryan County Oklahoma last summer.

Each year, Oklahoma growers harvest ca. 250,000 -300,000 acres of grain sorghum. In 2013, approximately 14 million bushels were harvested, worth about \$40 million. Preliminary results of some efficacy tests conducted in Texas and Louisiana indicate that the currently registered products (malathion, chlorpyrifos and dimethoate) provide inconsistent control. Pyrethroid insecticides registered for control are also ineffective. The listed products are also very broad-spectrum in their activity and thus are very hard on potential natural enemies. This presents the possibility of causing secondary pest outbreaks, such as with spidermites, or recurrent outbreaks of aphids.

This aphid was found in Bryan County Oklahoma last summer. We do not know the potential for this aphid to spread and cause major damage to sorghum production in Oklahoma. It appears to be a new 'biotype' that has recently switched to sorghum as a preferred host. We are concerned that this aphid caused such significant yield loss in Texas and Louisiana (40-50% loss) with apparently diminished ability to correct a severe outbreak with currently registered insecticides.

I am in full support of this Section 18 application which is also supported by the National Sorghum Producers as well as the Oklahoma Sorghum Association.

Sincerely,

A handwritten signature in cursive script that reads 'Tom A. Royer'.

Tom A. Royer
Extension Entomologist and IPM Coordinator





Dow AgroSciences

Dow AgroSciences LLC

9330 Zionsville Road Indianapolis, IN 46268 USA

www.dowagro.com

March 31, 2014

Ryan Williams
Oklahoma Department of Agriculture, Food & Forestry
Certification & Training Administrator
2800 N. Lincoln Blvd.
Oklahoma City, OK 73105

Re: Support letter for TransformTM WG Section 18 on sorghum

Dear Mr. Williams,

Per your request, this letter is to confirm that Dow AgroSciences supports the pursuit of a Section 18 emergency exemption for Transform WG to control sugarcane aphid in sorghum in the state of Oklahoma. Transform WG was registered by the US Environmental Protection Agency to control aphids and other pests on a number of crops in 2013. A tolerance petition was recently submitted to EPA in pursuit of a Section 3 registration in sorghum but that registration is not expected prior to the 2015 use season.

Transform WG provides excellent efficacy against aphids and the active ingredient, sulfoxaflor, represents a new class of chemistry with a novel mode of action. As such it controls pests resistant to other classes of chemistry, among other benefits.

If you have questions, please do not hesitate to contact me.

Sincerely,

A handwritten signature in black ink, appearing to read "Jamey Thomas".

Jamey Thomas, Ph.D.
US Regulatory Manager
Dow AgroSciences

cc: Tami Jones-Jefferson, DAS

TMTrademark of Dow AgroSciences LLC

2014 FIFRA SECTION 18

General information requirements of §40 CFR 166.20(a) in an application for a specific exemption.

TYPE OF EXEMPTION BEING REQUESTED

✓ SPECIFIC

QUARANTINE

PUBLIC HEALTH

SECTION 166.20(a)(1): IDENTITY OF CONTACT PERSONS

- i. This application to the Administrator of the Environmental Protection Agency (EPA) for a specific exemption to authorize the use of Sulfoxaflor (Transform® WG Insecticide EPA Reg. No. 62719-625) to control the newly introduced sugarcane aphid, *Melanaphis sp.* (thought to be *Melanaphis sacchari*) in sorghum by the Oklahoma Department of Agriculture, Food, & Forestry. Any questions related to this request should be addressed to:

Ryan Williams
Oklahoma Department of Agriculture, Food, & Forestry
Pesticide Program Administrator
2800 N. Lincoln Blvd.
Oklahoma City, Ok
Phone: (405) 522-5993
Fax: (405) 522-5986
Email: ryan.williams@ag.ok.gov

- ii. The following qualified experts are also available to answer questions:

University Representatives:

Tom Royer, PhD
IPM Coordinator
Oklahoma State University
127 NRC
Stillwater, Ok 74078
405-744-9406
tom.royer@okstate.edu

Registrant Representative:

Tami Jones-Jefferson

U.S. Regulatory Leader

U.S. Regulatory & Government Affairs - Crop Protection

Dow AgroSciences

9330 Zionsville Road

Indianapolis IN 46268

phone: 317.337.3574

email: tjjonesjefferson@dow.com

SECTION 166.20(a)(2): DESCRIPTION OF THE PESTICIDE REQUESTED

- i. **Common Chemical Name (Active Ingredient):** Sulfoxaflor

Trade Name and EPA Reg. No.: Transform® WG Insecticide, EPA Reg. No. 62719-625

Formulation: Active Ingredient 50%

SECTION 166.20(a)(3): DESCRIPTION OF THE PROPOSED USE

- i. **Sites to be treated:**
Sorghum fields (grain and forage) with the newly introduced sugarcane aphid, *Melanaphis sp.* statewide.
- ii. **Method of Application:**
Applications will be made by foliar application when populations reach economic threshold values.
- iii. **Rate of Application:**
0.75 – 1.5 oz of Transform® WG/acre (0.023 – 0.047 lb ai/acre)
- iv. **Maximum Number of Applications:**
2 applications per year (maximum of 3 oz/acre (0.094 lb ai/acre)
- v. **Total Acreage to be Treated:**
According to the Oklahoma State University 250,000-300,000 acres of sorghum is planted in Oklahoma annually.

vi. Total Amount of Pesticide to be used:

According to the previously mentioned statistics, if all 300,000 acres of sorghum were treated with the maximum rate (1.5 oz/acre or 0.047 lb ai/acre) and the maximum numbers of applications are made (2 applications or 3.0 oz/acre or 0.094 lb ai/acre) then 7,032 gallons of Transform® WG would be used in 2014.

vii. Restrictions and Requirements:

- Preharvest Interval: Do not apply within 7 days of harvest for grain or 14 days of harvest for forage or stover.
- Minimum Treatment Interval: Do not make applications less than 14 days apart.
- Do not make more than two applications per acre per year.
- Do not apply more than a total of 3.0 oz of Transform WG (0.09 lb ai of sulfoxaflor) per acre per year.

Duration of the Proposed use:

Spring through late summer

viii. Earliest Possible Harvest Date:

August 1st

SECTION 166.20(a)(4): ALTERNATIVE METHODS OF CONTROL

Registered Alternative Pesticides:

Of the registered alternative pesticides, only Dimethoate 4 EC (dimethoate, EPA Reg. No. 19713-231) has provided adequate control. Dimethoate is an organophosphate which is labeled for use on sorghum at 1 pint per acre. Dimethoate, which is highly toxic to bees, has a use restriction that does not allow its use during pollen shed in sorghum. Insects have historically shown resistance to organophosphates. Three other pesticides registered for use in sorghum did not provide adequate control of the aphid. Those pesticides are:

Karate® with Zeon™ Technology (Lambda Cyhalothrin 22.8%, EPA Reg. No. 100-1097)

Lorsban® Advanced, others (Chlorpyrifos 40.2%, EPA Reg. No. 62719-591)

Asana® XL (Esfenvalerate 8.4%, EPA Reg. No. 352-515)

Of the above mentioned insecticides, Karate® and Asana® are pyrethroids and Lorsban® is an organophosphate. Both pyrethroids and organophosphates have shown resistance potential. In field tests conducted in 2013 by Texas A&M AgriLife professionals, Karate® and Asana® both provided some initial population reduction when used at labeled rates. However, population spikes were observed soon after treatments in some instances. Chlorpyrifos did not provide satisfactory control at labeled rates.

A few varieties of resistant sorghum have been identified by researchers, but sufficient quantities of agronomically acceptable cultivars will not be available for the 2014 planting season.

SECTION 166.20(a)(5): EFFICACY OF USE PROPOSED UNDER SECTION 18

Two replicated field trials were conducted on the aphid in 2013. The first was conducted by Dr. Mo Way in August in China, Texas. The second replicated test was conducted in Weslaco, Texas, by Dr. Raul Villanueva and D. Sekula. In both trials, data showed that Transform® WG at 0.75/acre provided good control of *Melanaphis sp.*

SECTION 166.20(a)(6): EXPECTED RESIDUES FOR FOOD USES

Michael Hare, Ph.D.

Acute Assessment

Food consumption information from the USDA 1994-1996 and 1998 Nationwide Continuing Surveys of Food Intake by Individuals (CSFII) and maximum residues from field trials rather than tolerance-level residue estimates were used. It was assumed that 100% of crops covered by the registration request are treated and maximum residue levels from field trials were used.

Drinking water. Two scenarios were modeled, use of sulfoxaflor on non-aquatic row and orchard crops and use of sulfoxaflor on watercress. For the non-aquatic crop scenario, based on the Pesticide Root Zone Model/Exposure Analysis Modeling System (PRZM/EXAMS) and Screening Concentration in Ground Water (SCI-GROW) models, the estimated drinking water concentrations (EDWCs) of sulfoxaflor for acute exposures are 26.4 ppb for surface water and 69.2 ppb for ground water. For chronic exposures, EDWCs are 13.5 ppb for surface water and 69.2 ppb for ground water. For chronic exposures for cancer assessments, EDWCs are 9.3 ppb for surface water and 69.2 ppb for ground water. For the watercress scenario, the EDWCs for surface water are 91.3 ppb after one application, 182.5 ppb after two applications and 273.8 ppb after three applications.

Dietary risk estimates using both sets of EDWCs are below levels of concern. The non-aquatic-crop EDWCs are more representative of the expected exposure profile for the majority of the population. Also, water concentration values are adjusted to take into account the source of the water; the relative amounts of parent sulfoxaflor, X11719474, and X11519540; and the relative liver toxicity of the metabolites as compared to the parent compound.

For acute dietary risk assessment of the general population, the groundwater EDWC is greater than the surface water EDWC and was used in the assessment. The residue profile in groundwater is 60.9 ppb X11719474 and 8.3 ppb X11519540 (totaling 69.2 ppb). Parent sulfoxaflor does not occur in groundwater. The regulatory toxicological endpoint is based on neurotoxicity.

For acute dietary risk assessment of females 13-49, the regulatory endpoint is attributable only to the parent compound; therefore, the surface water EDWC of 9.4 ppb was used for this assessment.

A tolerance of 0.3 ppm for sulfoxaflor on grain sorghum has been established. There is no expectation of residues of sulfoxaflor and its metabolites in animal commodities as a result of the proposed use on sorghum. Thus, animal feeding studies are not needed, and tolerances need not be established for meat, milk, poultry, and eggs.

Drinking water exposures are the driver in the dietary assessment accounting for 100% of the exposures. Exposures through food (sorghum grain and syrup) are zero.

The acute dietary exposure from food and water to sulfoxaflor is 16% of the aPAD for children 1-2 years old and females 13-49 years old, the population groups receiving the greatest exposure.

Chronic Assessment

The same refinements as those used for the acute exposure assessment were used, with two exceptions: (1) average residue levels from crop field trials were used rather than maximum values and (2) average residues from feeding studies, rather than maximum values, were used to derive residue estimates for livestock commodities. It was assumed that 100% of crops are treated and average residue levels from field trials were used.

For chronic dietary risk assessment, the toxicological endpoint is liver effects, for which it is possible to account for the relative toxicities of X11719474 and X11519540 as compared to sulfoxaflor. The groundwater EDWC is greater than the surface water EDWC. The residue profile in groundwater is 60.9 ppb X11719474 and 8.3 ppb X11519540. Adjusting for the relative toxicity results in 18.3 ppb equivalents of X11719474 and 83 ppb X11519540 (totaling 101.3 ppb). The adjusted groundwater EDWC is greater than the surface water EDWC (9.3 ppb) and was used to assess the chronic dietary exposure scenario.

The maximum dietary residue intake via consumption of sorghum commodities would be only a small portion of the RfD (<0.001%) and therefore, should not cause any additional risk to humans via chronic dietary exposure. Consumption of sorghum by sensitive sub-populations such as children and non-nursing infants is essentially zero. Thus, the risk of these subpopulations to chronic dietary exposure to sulfoxaflor used on grain sorghum would be insignificant.

The major contributor to the risk was water (100%). There was no contribution from grain sorghum to the dietary exposure. All other populations under the chronic assessment show risk estimates that are below levels of concern.

Chronic exposure to sulfoxaflor from food and water is 18% of the cPAD for infants, the population group receiving the greatest exposure. There are no residential uses for sulfoxaflor.

Short-term risk. Because there is no short-term residential exposure and chronic dietary exposure has already been assessed, no further assessment of short-term risk is necessary, the chronic dietary risk assessment for evaluating short-term risk for sulfoxaflor is sufficient.

Intermediate-term risk. Intermediate-term risk is assessed based on intermediate-term residential exposure plus chronic dietary exposure. Because there is no residential exposure and chronic

dietary exposure has already been assessed, no further assessment of intermediate-term risk is necessary.

Cumulative effects. Sulfoxaflor does not share a common mechanism of toxicity with any other substances, and does not produce a toxic metabolite produced by other substances. Thus, sulfoxaflor does not have a common mechanism of toxicity with other substances.

Cancer. A nonlinear RfD approach is appropriate for assessing cancer risk to sulfoxaflor. This approach will account for all chronic toxicity, including carcinogenicity that could result from exposure to sulfoxaflor. Chronic dietary risk estimates are below levels of concern; therefore, cancer risk is also below levels of concern.

There is a reasonable certainty that no harm will result to the general population, or to infants and children from aggregate exposure to sulfoxaflor as used in this emergency exemption request.

SECTION 166.20(a)(7): DISCUSSION OF RISK INFORMATION

Human Health Effects – Michael Hare, Ph.D.

Ecological Effects – David Villarreal, Ph.D.

Environmental Fate – David Villarreal, Ph.D.

Human Health

Toxicological Profile

Sulfoxaflor is a member of a new class of insecticides, the sulfoximines. It is an activator of the nicotinic acetylcholine receptor (nAChR) in insects and, to a lesser degree, mammals. The nervous system and liver are the target organs, resulting in developmental toxicity and hepatotoxicity.

Developmental toxicity was observed in rats only. Sulfoxaflor produced skeletal abnormalities likely resulting from skeletal muscle contraction due to activation of the skeletal muscle nAChR in utero. Contraction of the diaphragm, also related to skeletal muscle nAChR activation, prevented normal breathing in neonates and increased mortality. The skeletal abnormalities occurred at high doses while decreased neonatal survival occurred at slightly lower levels.

Sulfoxaflor and its major metabolites produced liver weight and enzyme changes, and tumors in subchronic, chronic and short-term studies. Hepatotoxicity occurred at lower doses in long-term studies compared to short-term studies.

Reproductive effects included an increase in Leydig cell tumors which were not treatment related due to the lack of dose response, the lack of statistical significance for the combined tumors, and the high background rates for this tumor type in F344 rats. The primary effects on male reproductive organs are secondary to the loss of normal testicular function due to the size of the Leydig Cell adenomas. The secondary effects to the male reproductive organs are also not

treatment related. It appears that rats are uniquely sensitive to these developmental effects and are unlikely to be relevant to humans.

Clinical indications of neurotoxicity were observed at the highest dose tested in the acute neurotoxicity study in rats. Decreased motor activity was also observed in the mid- and high-dose groups. Since the neurotoxicity was observed only at a very high dose and many of the effects are not consistent with the perturbation of the nicotinic receptor system, it is unlikely that these effects are due to activation of the nAChR.

Tumors have been observed in rat and mouse studies. In rats, there were significant increases in hepatocellular adenomas in the high-dose males. In mice, there were significant increases in hepatocellular adenomas and carcinomas in high dose males. In female mice, there was an increase in carcinomas at the high dose. Liver tumors in mice were treatment-related. Leydig cell tumors were also observed in the high-dose group of male rats, but were not related to treatment. There was also a significant increase in preputial gland tumors in male rats in the high-dose group. Given that the liver tumors are produced by a non-linear mechanism, the Leydig cell tumors were not treatment-related, and the preputial gland tumors only occurred at the high dose in one sex of one species, the evidence of carcinogenicity was weak.

Ecological Toxicity

Sulfoxaflor (N-[methyloxido[1-[6-(trifluoromethyl)-3-pyridinyl]ethyl]-lambda 4-sulfanylidene]) is a new variety of insecticide as a member of the sulfoxamine subclass of neonicotinoid insecticides. It is considered an agonist of the nicotinic acetylcholine receptor and exhibits excitatory responses including tremors, followed by paralysis and mortality in target insects. Sulfoxaflor consists of two diastereomers in a ratio of approximately 50:50 with each diastereomer consisting of two enantiomers. Sulfoxaflor is systemically distributed in plants when applied. The chemical acts through both contact action and ingestion and provides both rapid knockdown (symptoms are typically observed within 1-2 hours of application) and residual control (generally provides from 7 to 21 days of residual control). Incident reports submitted to EPA since approximately 1994 have been tracked via the Incident Data System. Over the 2012 growing season, a Section 18 emergency use was granted for application of sulfoxaflor to cotton in four states (MS, LA, AR, TN). No incident reports have been received in association with the use of sulfoxaflor in this situation.

Sulfoxaflor is classified as practically non-toxic on an acute exposure basis, with 96-h LC₅₀ values of >400 mg a.i./L for all three freshwater fish species tested (bluegill, rainbow trout, and common carp). Mortality was 5% or less at the highest test treatments in each of these studies. Treatment-related sublethal effects included discoloration at the highest treatment concentration (100% of fish at 400 mg a.i./L for bluegill) and fish swimming on the bottom (1 fish at 400 mg a.i./L for rainbow trout). No other treatment-related sublethal effects were reported. For an estuarine/marine sheepshead minnow, sulfoxaflor was also practically non-toxic with an LC₅₀ of 288 mg a.i./L. Sublethal effects included loss of equilibrium or lying on the bottom of aquaria at 200 and 400 mg a.i./L. The primary degradate of sulfoxaflor is also classified as practically non-toxic to rainbow trout on an acute exposure basis (96-h LC₅₀ >500 mg a.i./L).

Adverse effects from chronic exposure to sulfoxaflor were examined with two fish species (fathead minnow and sheepshead minnow) during early life stage toxicity tests. For fathead minnow, the 30-d NOAEC is 5 mg a.i./L based on a 30% reduction in mean fish weight relative to controls at the next highest concentration (LOAEC=10 mg a.i./L). No statistically significant and/or treatment-related effects were reported for hatching success, fry survival and length. For sheepshead minnow, the 30-d NOAEC is 1.3 mg a.i./L based on a statistically significant reduction in mean length (3% relative to controls) at 2.5 mg a.i./L. No statistically significant and/or treatment-related effects were reported for hatching success, fry survival and mean weight.

The acute toxicity of sulfoxaflor was evaluated for one freshwater invertebrate species, the water flea and two saltwater species (mysis shrimp and Eastern oyster). For the water flea, the 48-h EC_{50} is >400 mg a.i./L, the highest concentration tested. For Eastern oyster, new shell growth was significantly reduced at 120 mg a.i./L (75% reduction relative to control). The 96-h EC_{50} for shell growth is 93 mg a.i./L. No mortality occurred at any test concentration. Mysid shrimp are the most acutely sensitive invertebrate species tested with sulfoxaflor based on water column only exposures, with a 96-h LC_{50} of 0.67 mg a.i./L. The primary degradate of sulfoxaflor is also classified as practically non-toxic to the water flea (EC_{50} >240 mg a.i./L).

The chronic effects of sulfoxaflor to the water flea were determined in a semi-static system over a period of 21 days to nominal concentrations of 6.25, 12.5, 25, 50 and 100 mg a.i./L. Adult mortality, reproduction rate (number of young), length of the surviving adults, and days to first brood were used to determine the toxicity endpoints. No treatment-related effects on adult mortality or adult length were observed. The reproduction rate and days to first brood were significantly ($p<0.05$) different in the 100 mg a.i./L test group (40% reduction in mean number of offspring; 35% increase in time to first brood). No significant effects were observed on survival, growth or reproduction at the lower test concentrations. The 21-day NOAEC and LOAEC were determined to be 50 and 100 mg a.i./L, respectively.

The chronic effects of sulfoxaflor to mysid shrimp were determined in a flow-through system over a period of 28 days to nominal concentrations of 0.063, 0.13, 0.25, 0.50 and 1.0 mg a.i./L. Mortality of parent (F_0) and first generation (F_1), reproduction rate of F_0 (number of young), length of the surviving F_0 and F_1 , and days to first brood by F_0 were used to determine the toxicity endpoints. Complete F_0 mortality (100%) was observed at the highest test concentration of 1.0 mg a.i./L within 7 days; no treatment-related effects on F_0/F_1 mortality, F_0 reproduction rate, or F_0/F_1 length were observed at the lower test concentrations. The 28-day NOAEC and LOAEC were determined to be 0.11 mg and 0.25 mg a.i./L, respectively.

Sulfoxaflor exhibited relatively low toxicity to aquatic non-vascular plants. The most sensitive aquatic nonvascular plant is the freshwater diatom with a 96-h EC_{50} of 81.2 mg a.i./L. Similarly, sulfoxaflor was not toxic to the freshwater vascular aquatic plant, *Lemna gibba*, up to the limit amount, as indicated by a 7-d EC_{50} for frond count, dry weight and growth rate of >100 mg a.i./L with no significant adverse effects on these endpoints observed at any treatment concentration.

Based on an acute oral LD_{50} of 676 mg a.i./kg bw for bobwhite quail, sulfoxaflor is considered slightly toxic to birds on an acute oral exposure basis. On a subacute, dietary exposure basis,

sulfoxaflor is classified as practically nontoxic to birds, with 5-d LC₅₀ values of >5620 mg/kg-diet for mallard ducks and bobwhite quail. The NOAEL from these studies is 5620 mg/kg-diet as no treatment related mortality or sublethal effects were observed at any treatment. Similarly, the primary degradate is classified as practically nontoxic to birds on an acute oral exposure basis with a LD₅₀ of >2250 mg a.i./kg bw. In two chronic, avian reproductive toxicity studies, the 20-week NOAELs ranged from 200 mg/kg-diet (mallard, highest concentration tested) to 1000 mg/kg-diet (bobwhite quail, highest concentration tested). No treatment-related adverse effects were observed at any test treatment in these studies.

For bees, sulfoxaflor is classified as very highly toxic with acute oral and contact LD₅₀ values of 0.05 and 0.13 µg a.i./bee, respectively, for adult honey bees. For larvae, a 7-d oral LD₅₀ of >0.2 µg a.i./bee was determined (45% mortality occurred at the highest treatment of 0.2 µg a.i./bee). The primary metabolite of sulfoxaflor is practically non-toxic to the honey bee. This lack of toxicity is consistent with the cyano-substituted neonicotinoids where similar cleavage of the cyanide group appears to eliminate their insecticidal activity. The acute oral toxicity of sulfoxaflor to adult bumble bees (*Bombus terrestris*) is similar to the honey bee; whereas its acute contact toxicity is about 20X less toxic for the bumble bee. Sulfoxaflor did not demonstrate substantial residual toxicity to honey bees exposed via treated and aged alfalfa (i.e., mortality was <15% at maximum application rates).

At the application rates used (3-67% of US maximum), the direct effects of sulfoxaflor on adult forager bee mortality, flight activity and the occurrence of behavioral abnormalities is relatively short-lived, lasting 3 days or less. Direct effects are considered those that result directly from interception of spray droplets or dermal contact with foliar residues. The direct effect of sulfoxaflor on these measures at the maximum application rate in the US is presently not known. When compared to control hives, the effect of sulfoxaflor on honey bee colony strength when applied at 3-32% of the US maximum proposed rate was not apparent in most cases. When compared to hives prior to pesticide application, sulfoxaflor applied to cotton foliage up to the maximum rate proposed in the US resulted in no discernible decline in mean colony strength by 17 days after the first application. Longer-term results were not available from this study nor were concurrent controls included. For managed bees, the primary exposure routes of concern include direct contact with spray droplets, dermal contact with foliar residues, and ingestion through consumption of contaminated pollen, nectar and associated processed food provisions. Exposure of hive bees via contaminated wax is also possible. Exposure of bees through contaminated drinking water is not expected to be nearly as important as exposure through direct contact or pollen and nectar.

In summary, sulfoxaflor is slightly toxic to practically non-toxic to fish and freshwater aquatic invertebrates on an acute exposure basis. It is also practically non-toxic to aquatic plants (vascular and non-vascular). Sulfoxaflor is highly toxic to saltwater invertebrates on an acute exposure basis. The high toxicity of sulfoxaflor to mysid shrimp and benthic aquatic insects relative to the water flea is consistent with the toxicity profile of other insecticides with similar MOAs. For birds and mammals, sulfoxaflor is classified as moderately toxic to practically non-toxic on an acute exposure basis. The threshold for chronic toxicity (NOAEL) to birds is 200 ppm and that for mammals is 100 ppm in the diet. Sulfoxaflor did not exhibit deleterious effects to terrestrial plants at or above its proposed maximum application rates.

For bees, sulfoxaflor is classified as very highly toxic. However, if this insecticide is strictly used as directed on the Section 18 supplemental label, no significant adverse effects are expected to Texas wildlife. Of course, standard precautions to avoid drift and runoff to waterways of the state are warranted. As stated on the Section 3 label, risk to managed bees and native pollinators from contact with pesticide spray or residues can be minimized when applications are made before 7 am or after 7 pm or when the temperature is below 55°F at the site of application.

Environmental Fate

Sulfoxaflor is a systemic insecticide which displays translaminar movement when applied to foliage. Movement of sulfoxaflor within the plant follows the direction of water transport within the plant (i.e., xylem mobile) as indicated by phosphorus translocation studies in several plants. Sulfoxaflor is characterized by a water solubility ranging from 550 to 1,380 ppm. Sulfoxaflor has a low potential for volatilization from dry and wet surfaces (vapor pressure = 1.9×10^{-8} torr and Henry's Law constant = 1.2×10^{-11} atm m³ mole⁻¹, respectively at 25 °C). Partitioning coefficient of sulfoxaflor from octanol to water (K_{ow} @ 20 °C & pH 7 = 6; Log K_{ow} = 0.802) suggests low potential for bioaccumulation. No fish bioconcentration study was provided due to the low K_{ow} , but sulfoxaflor is not expected to bioaccumulate in aquatic systems. Furthermore, sulfoxaflor is not expected to partition into the sediment due to low K_{oc} (7-74 mL/g).

Registrants tests indicate that hydrolysis, and both aqueous and soil photolysis are not expected to be important in sulfoxaflor dissipation in the natural environment. In a hydrolysis study, the parent was shown to be stable in acidic/neutral/alkaline sterilized aqueous buffered solutions (pH values of 5, 7 and 9). In addition, parent chemical as well as its major degradate, were shown to degrade relatively slowly by aqueous photolysis in sterile and natural pond water ($t^{1/2}$ = 261 to >1,000 days). Furthermore, sulfoxaflor was stable to photolysis on soil surfaces. Sulfoxaflor is expected to biodegrade rapidly in aerobic soil (half-lives <1 day). Under aerobic aquatic conditions, biodegradation proceeded at a more moderate rate with half-lives ranging from 37 to 88 days. Under anaerobic soil conditions, the parent compound was metabolized with half-lives of 113 to 120 days while under anaerobic aquatic conditions the chemical was more persistent with half-lives of 103 to 382 days. In contrast to its short-lived parent, the major degradate is expected to be more persistent than its parent in aerobic/anaerobic aquatic systems and some aerobic soils. In other soils, less persistence is expected due to mineralization to CO₂ or the formation of other minor degradates.

In field studies, sulfoxaflor has shown similar vulnerability to aerobic bio-degradation in nine out of ten terrestrial field dissipation studies on bare-ground/cropped plots (half-lives were <2 days in nine cropped/bare soils in CA, FL, ND, ON and TX and was 8 days in one bare ground soil in TX). The chemical can be characterized by very high to high mobility (K_{foc} ranged from 11-72 mL g⁻¹). Rapid soil degradation is expected to limit chemical amounts that may potentially leach and contaminate ground water. Contamination of groundwater by sulfoxaflor will only be expected when excessive rain occurs within a short period (few days) of multiple applications in vulnerable sandy soils. Contamination of surface water by sulfoxaflor is expected to be mainly related to drift and very little due to run-off. This is because drifted sulfoxaflor that reaches aquatic systems is expected to persist while that reaching the soil system is expected to degrade quickly with slight chance for it to run-off.

When sulfoxaflor is applied foliarly on growing crops it is intercepted by the crop canopy. Data presented above appear to indicate that sulfoxaflor enters the plant and is incorporated in the plant foliage with only limited degradation. It appears that this is the main source of the insecticide sulfoxaflor that would kill sap sucking insects. This is because washed-off sulfoxaflor, that reaches the soil system, is expected to degrade.

In summary, sulfoxaflor has a low potential for volatilization from dry and wet surfaces. This chemical is characterized by a relatively higher water solubility. Partitioning coefficient of sulfoxaflor from octanol to water suggests low potential for bioaccumulation in aquatic organisms such as fish. Sulfoxaflor is resistant to hydrolysis and photolysis but transforms quickly in soils. In contrast, sulfoxaflor reaching aquatic systems by drift is expected to degrade rather slowly. Partitioning of sulfoxaflor to air is not expected to be important due to the low vapor pressure and Henry's Law constant for sulfoxaflor. Exposure in surface water results from drifted parent as only minor amounts is expected to run-off only when rainfall and/or irrigation immediately follow application. The use of this insecticide is not expected to significantly adversely impact Texas ecosystems with use according to the Section 18 label with this application. Of course, caution is needed to prevent exposure to water systems because of toxicity issues to aquatic invertebrates. As stated on the Section 3 label, this product should never be applied directly to water, to areas where surface water is present or to intertidal areas below the mean water mark. Do not contaminate water when disposing of equipment rinsates.

Endangered and Threatened Species in Texas

No impacts are expected on endangered and threatened species by this very limited use of this insecticide as delineated in the Section 18 application. Sulfoxaflor demonstrates a very favorable ecotoxicity and fate profile as stated above and should not directly impact any protected mammal, fish, avian, or plant species. This product does adversely affect insects and aquatic invertebrates, especially bees, but the limited exposure to these species should not negatively affect endangered and threatened species in Oklahoma. As always, the label precautions need be strictly adhered to.

SECTION 166.20(a)(8): COORDINATION WITH OTHER AFFECTED STATE OR FEDERAL AGENCIES

The following state/federal agencies were notified of the Oklahoma Department of Agriculture, Food, and Forestry actions to submit an application for a specific exemption to EPA:

- Oklahoma Department of Environmental Quality (ODEQ), Air Quality Control
- Oklahoma Department of Environmental Quality (ODEQ), Water Quality
- Oklahoma Department of Health
- Oklahoma Department of Wildlife Conservation
- U.S. Fish and Wildlife Department

Responses from these agencies will be forwarded to EPA immediately if and when received by ODA.

SECTION 166.20(a)(9): ACKNOWLEDGEMENT BY THE REGISTRANT

Dow AgroScience has been notified of this agency's intent regarding this application (see attached letter of support). They have also provided a copy of a label with the use directions for this use (although this use is dependent upon the approval of this section-18 by EPA).

SECTION 166.20(a)(10): DESCRIPTION OF PROPOSED ENFORCEMENT PROGRAM

The State Legislature has endowed the ODAFF with the authority to regulate the distribution, storage, sale, use and disposal of pesticides in the state of Oklahoma. In addition, the EPA/ODAFF grant enforcement agreement provides the Department with the authority to enforce the provisions of the FIFRA, as amended, within the state. Therefore, the Department is not lacking in authority to enforce the provisions of an EPA approved specific exemption. If this specific exemption request is approved, ODAFF Pesticide Enforcement Specialists will make a number of random, unannounced calls on both growers and applicators to check for compliance with provisions of the specific exemption. If violations are discovered appropriate enforcement will be taken.

SECTION 166.20(a)(11): REPEAT USES

This is the first time Oklahoma Department of Agriculture, Food, & Forestry has applied for this specific exemption.

SECTION 166.20(b)(1): NAME OF THE PEST

Melanaphis sp. (thought to be *Melanaphis sacchari*)

SECTION 166.20(b)(2): DISCUSSION OF EVENTS OR CIRCUMSTANCES WHICH BROUGHT ABOUT THE EMERGENCY SITUATION

Oklahoma producers generally produces somewhere in the neighborhood of 250,000 -300,000 acres of grain sorghum annually. Preliminary results of some efficacy tests conducted in Texas and Louisiana indicate that the currently registered products malathion, chlorpyrifos and dimethoate provide inconsistent control. Pyrethroid insecticides registered for control are also ineffective. The listed products are also very broad-spectrum in their activity, being very hard on potential natural enemies. This presents the possibility of causing secondary pest outbreaks, such as with spidermites, or recurrent outbreaks of aphids.

This aphid was found in Bryan County Oklahoma last summer. We do not know the potential for this aphid to spread and cause major damage to sorghum production in Oklahoma. It appears

to be a new 'biotype' that has recently switched to sorghum as a preferred host. We are concerned that this aphid caused such significant yield loss in Texas and Louisiana (40-50% loss) with apparently diminished ability to correct a severe outbreak with currently registered insecticides.

**SECTION 166.20(b)(3): DISCUSSION OF ANTICIPATED RISKS TO
ENDANGERED OR THREATENED SPECIES, BENEFICIAL ORGANISMS, OR
THE ENVIRONMENT**

As discussed previously, it is not anticipated that there should be any anticipated risks to endangered or threatened species, beneficial organisms or the environment if the application is made according to the section 18 use directions.

SECTION 166.20(b)(4): DISCUSSION OF SIGNIFICANT ECONOMIC LOSS

As mentioned above this pest was found in Southern Oklahoma along the Texas border last year, and we do not know the potential for it to spread. However, growers widely reported 20 to 50% yield loss in infested fields. Scouts have observed the aphid successfully overwintering in volunteer sorghum in the South Texas region, increasing the probability of a more widespread outbreak in 2014. Dr. Mo Way reported one producer in Chambers County did not harvest his grain sorghum because the aphid damage was so severe. He also reported a producer in Liberty County suffered a yield loss of 50% in an infested field as compared to another field where the aphid did not damage the crop. Dr. Raul Villanueva reported two seed increase plots at the Texas A&M AgriLife Center in Weslaco were completely lost due to the aphid. Dr. Villanueva has also received information from Mexico that research plots at the Rio Bravo Agricultural Station were all devastated by this aphid and the growers had up to 60% loss in San Fernando and Ciudad Victoria. These locations are 2 and 4 hours from the Texas border respectively.



2013 Outbreak of Sorghum/Sugarcane Aphid on Sorghum

Michael Brewer

Texas A&M AgriLife Research
Department of Entomology
Corpus Christi

TEXAS A&M
AGRI LIFE
RESEARCH

Outbreak of Sorghum/Sugarcane Aphid on Sorghum:
First Detections, Distribution, and Notes on Management

TEXAS A&M
AGRI LIFE
EXTENSION

M. Brewer^{1,4}, M. Way^{1,4}, S. Armstrong², S. Biles^{3,4}, D. Sekula^{3,4}, J. Swart^{3,4}, C. Crumley^{3,4}, A. Knutson^{3,4}, R. Villanueva^{3,4}, R. Parker^{3,4}, G. Odvody¹, and D. Ragsdale⁴

¹ Texas A&M AgriLife Research, Corpus Christi and Beaumont; ² USDA ARS, Stillwater, OK;

³ Texas A&M AgriLife Extension, Port Lavaca, Weslaco, Commerce, Wharton, Dallas, and Corpus Christi; ⁴ Texas A&M University, Department of Entomology

<http://ccag.tamu.edu/entomology/>

Outline

- I. Identification, Biology, Crop Hosts, and Distribution**
- II. Sorghum Damage and Harvest Problems**
- III. Overwintering Potential along the Gulf Coast**
- IV. Observations on Plant Resistance and Natural Enemies**
- V. Insecticide Efficacy**
- VI. Management Approaches in 2014**
- VII. Research & Extension Activities in 2014**



M. Brewer, AgriLife Research



R. Villanueva, AgriLife Extension

I. Identification, Biology, Crop Hosts, and Distribution

**Grass hosts where nymphs & adults were observed:
Sorghum, johnson grass,
Sorghum-sudan (Hay grazer)**

**Underside of leaves & stems,
Bottom to top of plant**

**No observations on
sugarcane**

**Observed on corn, but no
reproduction**

Sorghum is a great SA host



Sugarcane (sorghum) aphid and others found on sorghum: 'stove pipes', no 'spots', no 'spines', color varies (darker buff in winter)

Corn leaf aphid

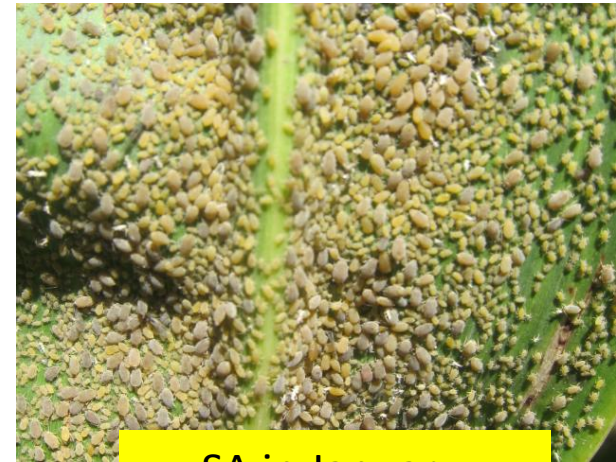


Yellow sugarcane aphid



SA in August, Corpus Christi

G. Odvody/M. Brewer, AgriLife Research



SA in January, remnant sorghum, RGV

R. Villanueva, AgriLife Extension

SA: sugarcane/sorghum aphid

Greenbug aphid

Wind-blown winged females establishes a colony

Mostly wingless asexual reproduction for rapid increase (clones)

Weekly Repro. Rate: ~8-20 nymphs per adult (live birth)

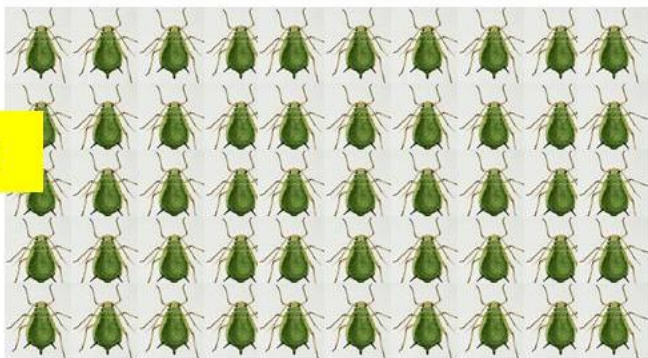
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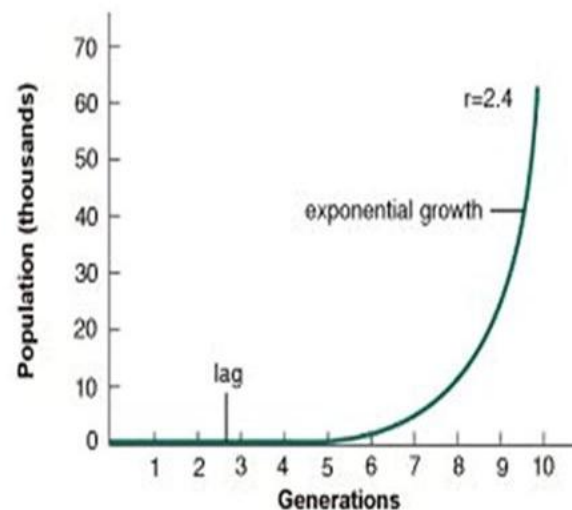
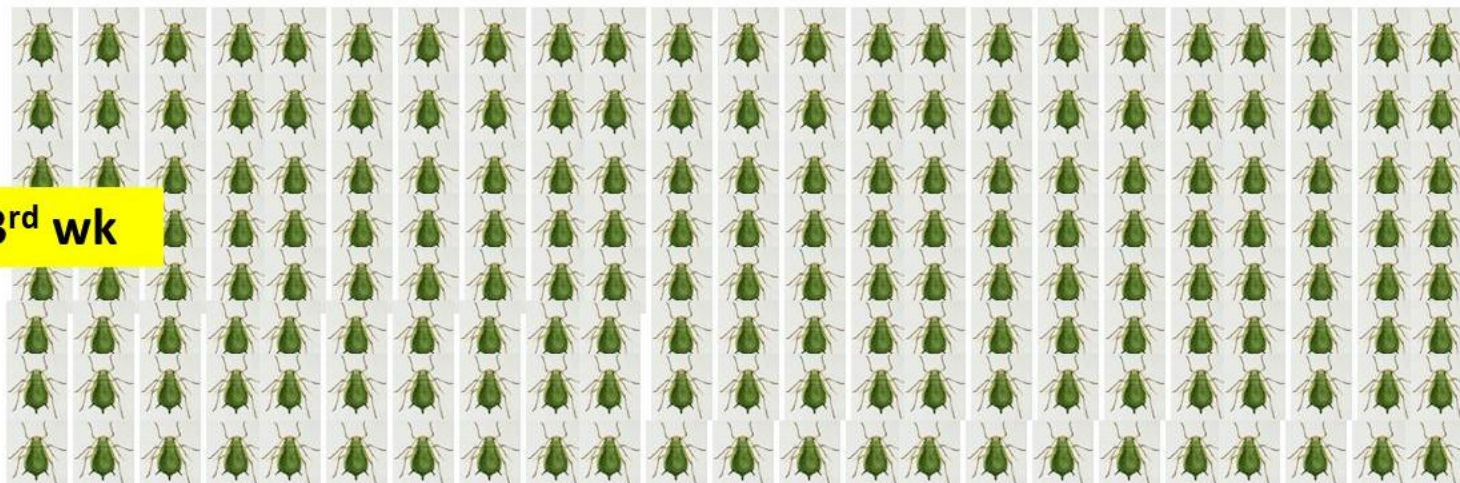
1st wk

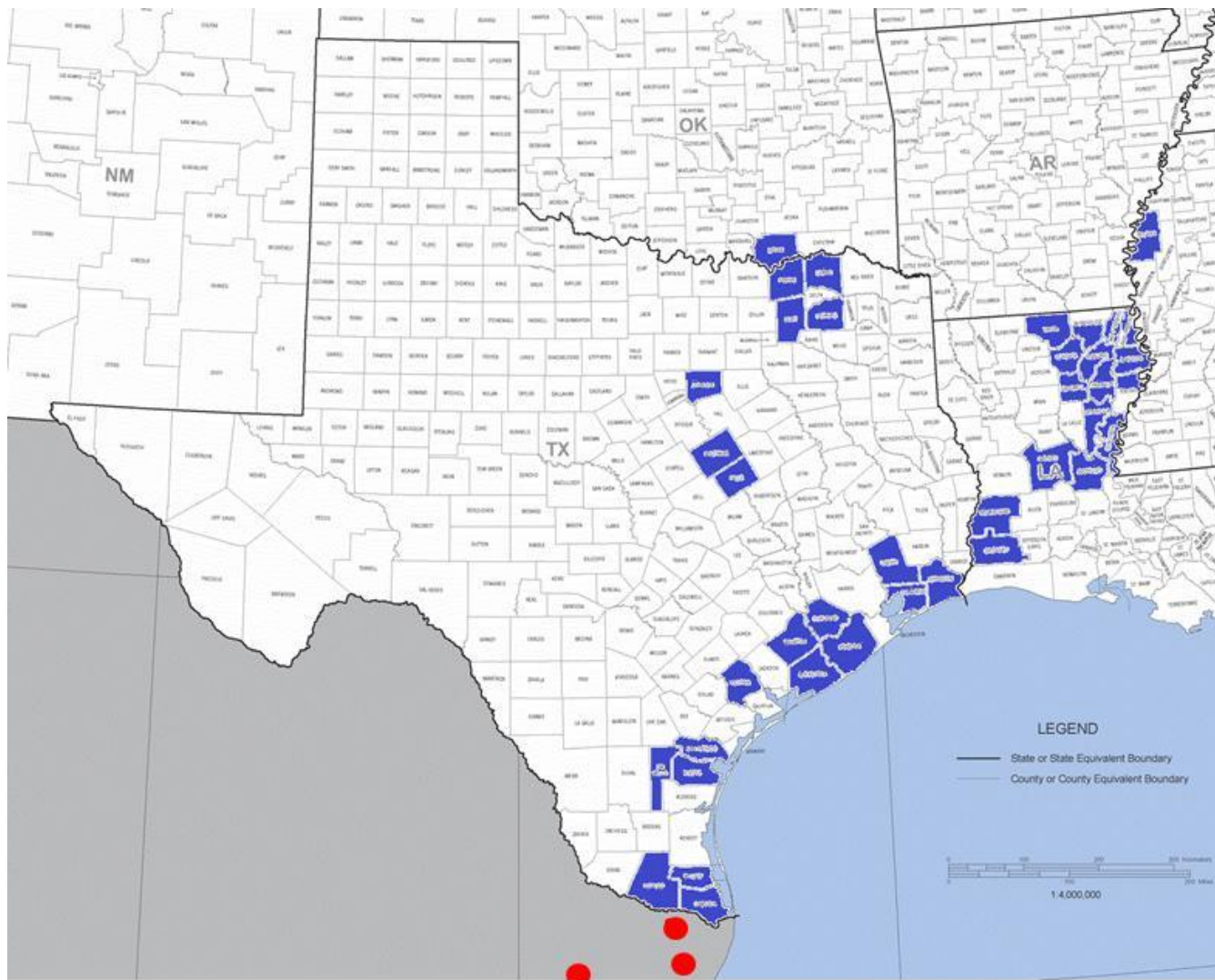


2nd wk



3rd wk





The new aphid pest of sorghum was detected in 38 counties and parishes of Texas, Louisiana, Oklahoma, and Mississippi, and one state in Mexico (red dots) in 2013 (very likely in Arkansas as well). It spread quickly across the region. All sorghum growing counties in the region likely at risk.

Courtesy of Melanaphis Task Force,
D. Anderson/R. Villanueva, AgriLife Research/Extension

II. Sorghum Damage and Harvest Problems



Outbreak SA of sorghum in summer 2013 (top left, Beaumont), fall population on Johnson grass (bottom left, Corpus Christi). Note the presence of a few winged aphids and many unwinged aphids. Severe whole plant damage (top right, Beaumont), and close-up of sooty mold/honeydew damage (bottom right, Corpus Christi).

Courtesy of M. Way/M. Brewer/
G. Odvody, AgriLife Research,
R. Villanueva, AgriLife Extension

Plant damage caused by general plant decline

Yield loss in 1st yr primarily by honeydew/sooty mold affecting harvest

No detection of acute plant toxicity or disease introduction (IPM can be successful)

Damage estimates, 2013 and projected (no control)

Core infestation zone: Gulf Coast of TX, LA, Mexico

~10% fields infested during 1st yr invasion, some areas higher (RGV)

25-50% yield loss: primary harvest problems due to honeydew/sooty mold and some direct plant damage, especially in south

Some total loss due to crop abandonment

X Possibility one: Outbreak fizzles, one year loss

Possibility two: 2014 similar to 2013,

Geographic expansion modest

Infestation rate (10%) and yield loss rate (20-50%) stays the same

\$8.00 per hundred weight, harvest potential 4,500 lbs/ac

→ up to \$50M statewide (W. Cleveland, TGSP)

Possibility three: invasion strengthens

Geographic expansion great, including unanticipated north & west

Severity increases across season (Texas Gulf Coast)!!

Infestation rate increase (50% of fields)

Yield loss rate moving toward 50%

→ up to \$250M statewide

R. Villanueva, AgriLife Extension



M. Brewer, AgriLife Research



M. Way, AgriLife Research



R. Villanueva, AgriLife Extension

**Underside of leaves & stems,
Bottom to top of plant**

**Commercial sorghums are very good hosts,
leaves, stems, and even head stalks**

III. Overwintering Potential along the Gulf Coast

G. Odvody/M. Brewer, AgriLife Research



SA found throughout Coastal Bend (summer expanding colony), new founding colony with one winged adult and first offspring (first onto sorghum, spring concern)



R. Villanueva, AgriLife Extension

SA found RGV, Corpus Christi, and Victoria areas (Nov.- Jan. overwintering), new founding colony with one winged adult and first offspring (fall & spring concern)

IV. Observations on Plant Resistance and Natural Enemies

Screening existing CVs and breeding material has begun

Texas A&M AgriLife

B. Rooney, G. Peterson

USDA ARS

S. Armstrong

Predatory beetles and flies, and parasites (black mummies) have been observed but not limiting aphid increase

M. Brewer, J. Woolley, M. Way, R. Villanueva, S. Biles



M. Brewer,
AgriLife Research

R. Villanueva,
AgriLife Extension



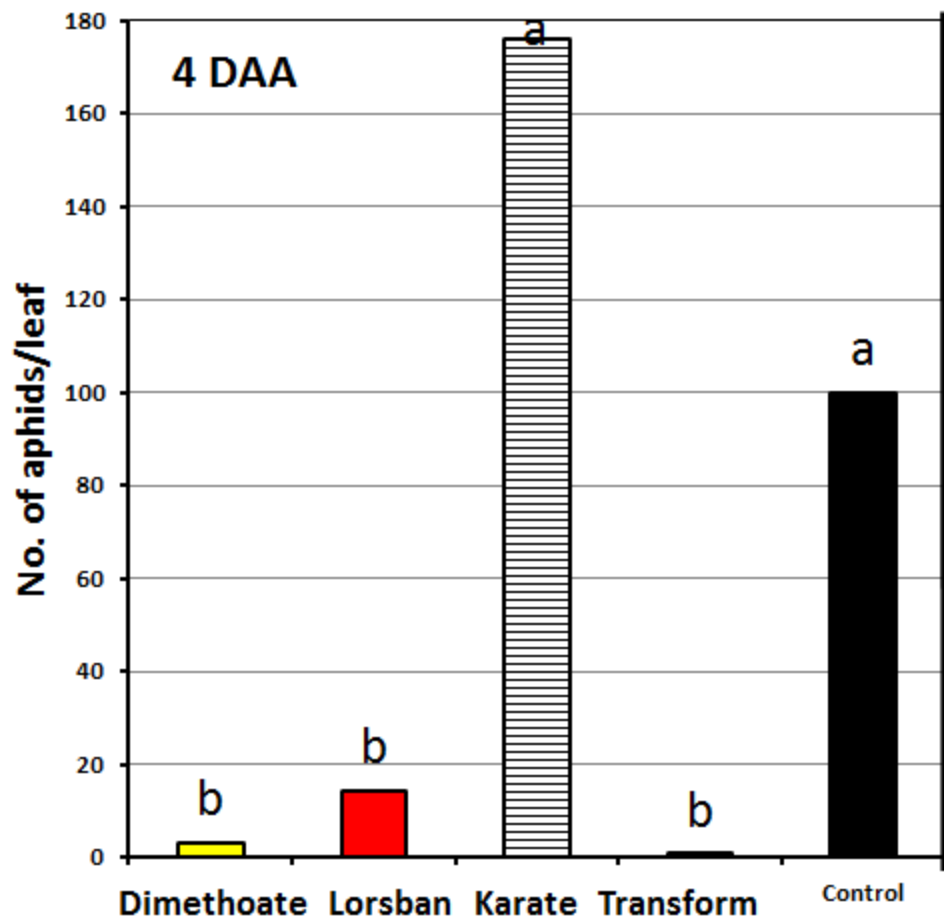
R. Villanueva,
AgriLife Extension



V. Insecticide Efficacy

Beaumont, M. Way, August 2013

Field previously had been treated with Lorsban 4E: 2 or 3 times



R. Villanueva, AgriLife Extension

Insecticide	Rate	LA	TX
Dimethoathe 4EC	1pt/A	✓	✓
Lorsban Advanced	1 qt/A	✓	✓
Karate Z	2 fl oz/A	✓	✗
Asana	10 oz/A	✗	✓
Transform WG	0.75 oz/A	✓	✓
Admire Pro	8 oz/A	✗	✓
Untreated Control	-	-	-

Additional studies by D. Sekula, AgriLife Extension,
and D. Kerns, LSU

Transform: not registered, Section 18 emergency use application (TX, LA)
Dimethoate: Used for other sorghum aphids

VI. Management Approaches in 2014

Monitoring: where and when to look and spray

Core infestation zone: Assume new sorghum re-infests earlier than 2013, if successful local overwintering

Preplant: Inspect remnant sorghum and johnson grass for winter survivors and winged aphids (wind-aided movement to newly planted fields)

R. Villanueva, AgriLife Extension



SA in January,
remnant sorghum,
RGV



SA new colony on
johnson grass

R. Villanueva, AgriLife Extension

Monitoring: where and when to look and spray

Emergence to head elongation: Inspect plant from bottom to top, underside of leaves, look for new colonies and expanding colonies
If colonies well established with honeydew, any seed treatment effects not lasting, good justification to spray, check following weeks

One head elongated, check for aphids on stalk and head



G. Odvody/M. Brewer, AgriLife Research

M. Way/M. Brewer, AgriLife Research

R. Villanueva, AgriLife Extension

Quiz: Aphid monitoring, correct id, and timely action important

S. Armstrong/Grantham, USDA ARS/OSU



G. Odvody/M. Brewer, AgriLife Research



R. Villanueva, AgriLife Extension

Which is SA? What are the others?

Which is a summer colony?

Corn leaf aphid



Yellow sugarcane aphid



G. Odvody/M. Brewer, AgriLife Research
Summer expanding colony,
Corpus Christi



SA: sugarcane/sorghum
aphid



Greenbug aphid



R. Villanueva, AgriLife Extension
Nov.- Jan. overwintering, RGV,
Corpus Christi, Victoria

Why is SA so special in South Texas?

Current sorghums grown are excellent hosts

Excellent reproduction, survival, honeydew production

Direct sorghum damage and harvest problems

Weather is great for semi-tropical aphid (need to ck overwintering)

Natural enemies seen but often lags behind aphids in initial invasion

Invasion front present various possibilities of continuing impact

Possibility two: 2014 similar to 2013,

Geographic expansion modest

Infestation rate (10%) and yield loss rate (20-50%)

→ up to \$50M statewide (W. Cleveland, TGSP)

Possibility three: invasion strengthens

Geographic expansion great

Severity increases across season (Texas Gulf Coast)!!

Infestation rate increase (50% of fields)

Yield loss rate moving toward 50%

→ up to \$250M statewide

VII. Research & Extension Activities in 2014

Survey fields for SA and natural enemies

Selected fields intensively in core infestation zone

Distribute results throughout TX, LA, OK

Insecticide efficacy (RGV, Lower and Upper Gulf Coast, LA)

Economic thresholds (RGV, Lower and Upper Gulf Coast, LA)

Sorghum germplasm screens (College Stn, Lubbock, ARS)

Natural enemies (College Station, Corpus Christi, ARS)

Presentations

Sorghum U (Robstown)

Rio Grande Valley Cotton & Grain Conference

Feed Grain & Cotton Conference (Wharton)

Crop Symposium Conference (Corpus Christi R&E Center)

OK, LA Producer Meetings

Entomological Society of America

Webinar, to be posted Feb. 7 at

<http://ccag.tamu.edu/faculty-staff/corpus-christi-center/>

Thanks: Melanaphis Task Force & many others



Outbreak of Sorghum/Sugarcane Aphid on Sorghum: First Detections, Distribution, and Notes on Management



M. Brewer^{1,4}, M. Way^{1,4}, S. Armstrong², S. Biles^{3,4}, D. Sekula^{3,4}, J. Swart^{3,4}, C. Crumley^{3,4}, A. Knutson^{3,4}, R. Villanueva^{3,4}, R. Parker^{3,4}, G. Odvody¹, and D. Ragsdale⁴

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G. Odvody, M. Brewer, D. Anderson, R. Villanueva, M. Way,
S. Armstrong, D. Sekula, and D. Kerns for slides and information
from Texas, Louisiana, Oklahoma

Texas A&M Dep. of Entomology, AgriLife Research & Extension

Texas Grain Sorghum Producers Board Research Grant

United Sorghum Checkoff

Industry for providing insecticide products

Texas Department of Agriculture

<http://ccag.tamu.edu/entomology/>

ELECTRONIC CODE OF FEDERAL REGULATIONS

e-CFR Data is current as of January 15, 2014

Title 40: Protection of Environment

PART 180—TOLERANCES AND EXEMPTIONS FOR PESTICIDE CHEMICAL RESIDUES IN FOOD

Subpart C—Specific Tolerances

§180.668 Sulfoxaflor; tolerances for residues.

(a) *General.* Tolerances are established for residues of the insecticide sulfoxaflor, including its metabolites and degradates, in or on the commodities in the table. Compliance with the tolerance levels specified is to be determined by measuring only sulfoxaflor (*N*-[methyloxydo[1-[6-(trifluoromethyl)-3-pyridinyl]ethyl]- γ^4 -sulfanylidene]cyanamide).

Commodity	Parts per million
Almond, hulls	6.0
Barley, grain	0.40
Barley, hay	1.0
Barley, straw	2.0
Bean, dry seed	0.20
Bean, succulent	4.0
Beet, sugar, dried pulp	0.07
Beet, sugar, molasses	0.25
Berry, low growing, subgroup 13-7G	0.70
Cattle, fat	0.10
Cattle, meat	0.15
Cattle, meat byproducts	0.40
Cauliflower	0.08
Citrus, dried pulp	3.6
Cotton, gin byproducts	6.0
Cotton, hulls	0.35
Cottonseed subgroup 20C	0.20
Fruit, citrus, group 10-10	0.70
Fruit, pome, group 11-10	0.50
Fruit, small, vine climbing, subgroup 13-07F, except fuzzy kiwi fruit	2.0
Fruit, stone, group 12	3.0
Goat, fat	0.10
Goat, meat	0.15
Goat, meat byproducts	0.40
Grain, aspirated fractions	20.0
Grape, raisin	6.0
Hog, fat	0.01

Hog, meat	0.01
Hog, meat byproducts	0.01
Horse, fat	0.10
Horse, meat	0.15
Horse, meat byproducts	0.40
Leafy greens, subgroup 4A	6.0
Leafy petiole, subgroup 4B	2.0
Milk	0.15
Nuts, tree, group 14	0.015
Onion, bulb, subgroup 3-07A	0.01
Onion, green, subgroup 3-07B	0.70
Pistachio	0.015
Poultry, eggs	0.01
Poultry, fat	0.01
Poultry, meat	0.01
Poultry, meat byproducts	0.01
Rapeseed, meal	0.50
Rapeseed subgroup 20A	0.40
Sheep, fat	0.10
Sheep, meat	0.15
Sheep, meat byproducts	0.40
Soybean, seed	0.20
Tomato, paste	2.60
Tomato, puree	1.20
Vegetable, <i>brassica</i> , leafy, group 5, except cauliflower	2.0
Vegetable, cucurbit, group 9	0.40
Vegetable, fruiting, group 8-10	0.70
Vegetable, leaves of root and tuber, group 2	3.0
Vegetable, legume, foliage, group 7	3.0
Vegetable, root and tuber, group 1	0.05
Watercress	6.0
Wheat, forage	1.0
Wheat, grain	0.08
Wheat, hay	1.5
Wheat, straw	2.0

(b) *Section 18 emergency exemptions.* [Reserved]

(c) *Tolerances with regional registrations.* [Reserved]

(d) *Indirect or inadvertent residues.* [Reserved]

[77 FR 59565, Sept. 28, 2012, as amended at 78 FR 38227, June 26, 2013]

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Dow AgroSciences

Dow AgroSciences LLC

9330 Zionsville Road

Indianapolis, IN 46268-1054 USA

Transform[®] WG

EPA Reg. No. 62719-625

Section 18 Specific Exemption

Control of Sugarcane Aphid (*Melanaphis sacchari*) in Sorghum (For Distribution and Use Only in Oklahoma)

- **This Specific Exemption is effective XXXX through XXXX.**
- This labeling must be in the possession of the user at the time of application.
- Read the label affixed to the container for Transform[®] WG insecticide before applying. Carefully follow all precautionary statements and applicable use directions.
- Use of Transform WG according to this supplemental labeling is subject to all use precautions and limitations imposed by the label affixed to the container for Transform WG.

Directions for Use

Pests and Application Rates:

Pests	Transform WG (oz/acre)
Sugarcane aphid	0.75 – 1.5 (0.023 – 0.047 lb ai/acre)

Application Timing: Treat in accordance with local economic thresholds. Consult your Dow AgroSciences representative, cooperative extension service, certified crop advisor or state agricultural experiment station for any additional local use recommendations for your area.

Application Rate: Use a higher rate in the rate range for heavy pest populations.

Restrictions:

- **Preharvest Interval:** Do not apply within 7 days of harvest for grain or 14 days of harvest for forage or stover.
- **Minimum Treatment Interval:** Do not make applications less than 14 days apart.
- Do not make more than two applications per acre per year.
- Do not apply more than a total of 3.0 oz of Transform WG (0.09 lb ai of sulfoxaflor) per acre per year.

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Approved: __/__/__

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